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HISTOCHEMICAL STUDY OF NON-SPECIFIC ESTERASES MOTOR UNIT OF THE RABBIT

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SUMMARY

The distribution of alpha naphthyl acetate esterase and naphthol AS-D acetate esterase was studied in the motor neurons of the ventral horn of the spinal cord segments L4, L5, and SI, supplying the extensor digitorum longus muscle (EDL) of the rabbit together with the peripheral nerve supplying EDL and the muscle itself.

Strong activity was demonstrated in the motor neurons, in schwann cells and in the axons of peripheral nerve bundles.

Abundant reaction was located in the motor end-plates of muscle fibres making them easily recognizable. The biological interpretation of esterases was discussed.

INTRODUCTION

Alpha naphthyl acetate esterase and naphthol AS-D acetate esterase belong to the family of carboxylic ester hydrolases (E.C.3.1.1), which include a variety of enzymes that probably possess different functional roles in vivo, but have the common link of having esters of carboxylic acids as substrates. These esterases include carboxylesterase, arylesterase, acetyesterase, often collectively termed non-specific esterases, and cholinesterases. Alpha acetate is cleaved by nearly all non-specific esterases while the acetate of naphthol AS-D is cleaved mainly by arylesterase and acetyesterase(1).

Esterases are widely distributed in mammalian tissues particularly the nervous tissue where cholinesterases have prime importance in cholinergic transmission, and they are most commonly invested of the carboxyl ester

hydrolases. Cholin esterase also demonstrated in the perikarya of motor neurons of spinal cord (2). In many tissues distribution and level of biochemical activity of esterases indicate a particular functional state. In addition there is also histochemical evidency of a relationship between esterases and function (3).

The present work studies the distribution and localization of alpha naphthyl acetate esterase and naphtho AS-D acetate esterase in the motor units of the extensor digitorum longus (EDL) muscle of the rabbit. This is performed by investigating the histochemical activity of these esterases in the motor neurons supplying extrafusal fibres (alpha motor neurones) of EDL which are arranged in cell defined columns through L4, L5 and S1 segments of the spinal cord (4).

The motor nerve to EDL was studied as well by investigating the intramuscular nerve bundles, since the number of alpha motor axon profiles can be up to 40 per cent greater than in sections cut more centrally, the motor nerve fibres supplying EDL appear as relatively large myelinated fibres approximately 2-20µm in diameter (5). Lastly the EDL is studied with particular attention to the motor end-plates.

MATERIALS AND METHODS

Twelve New Zealand white rabbit (*Lepus europaeus*) weighing 400-900g. were used for investigation, each was anaesthetized by ether then exsanguinated. The EDL of both legs were dissected out. The motor nerve to EDL was studied by investigating the intramuscular nerve bundles since the number of alpha motor axon profiles can be up to 40 per cent greater than in sections cut more centrally (5). The spinal representation of EDL according to Sherrard (4) is in cell columns at L4, L5, and S1 segmental levels, noting that to muscles of lower limbs, flexor muscles are supplied by columns situated medial and caudal to those supplying corresponding extensor muscles.

The tissues were rapidly frozen in liquid nitrogen (-196 °C), serial sections, 8u each, were cut out by cryostat (Slee), then fixed in formalin vapour.

The histochemical demonstration of alpha naphthyl acetate esterase and naphthol AS-D acetate esterase were done by the technique of simultaneous azo-coupling method using the naphthol esters (alpha naphthyl acetate and naphthol AS-D acetate) as substrates, and hexazotized pararosaniline and fast blue RR as coupling agents respectively (6).

RESULTS

In the spinal cord, marked activity for non-specific esterase is demonstrated in the perikaryons of neurons of the ventral horn, both in large alpha motor neurons and smaller internuncial neurons (Fig.1), a strong reaction for non-specific esterases is seen in the grey matter (Fig.2).

Alpha naphthyl acetate esterase produce a strong dotlike reaction in schwann cells of nerve bundles with a lot of diffusion, no reaction is demonstrated in the axons (Fig.3).

Naphthol AS-D acetate esterase produces more localized and multiple granular reaction in nerve bundles with diffuse reaction in the axons (Fig.4).

Both esterases are abundant at the motor end-plates of the muscle fibres making them easily recognizable (Fig.5 & 6).

DISCUSSION

This study has shown that alpha naphthyl acetate esterase and naphthol AS-D acetate esterase are distributed throughout the motor unit, from the neurons of the spinal cord down to the muscle.

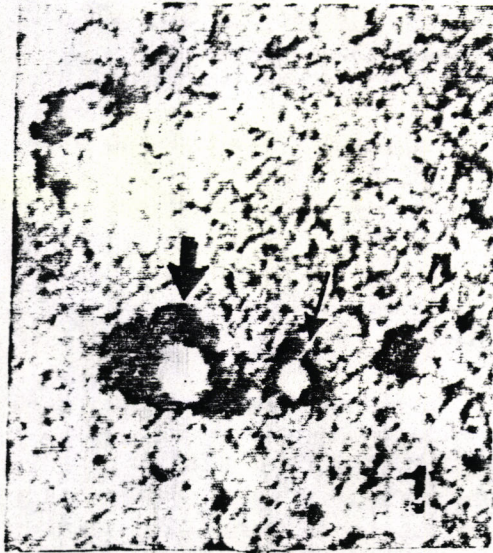


Fig. (1): Alpha naphthyl acetate esterase in the alpha motor neuron (thick arrow) and internuncial neuron (thin arrow) of the anterior horn spinal cord.



Fig. (2): Non-specific esterases reaction in the anterior horn of the spinal cord.

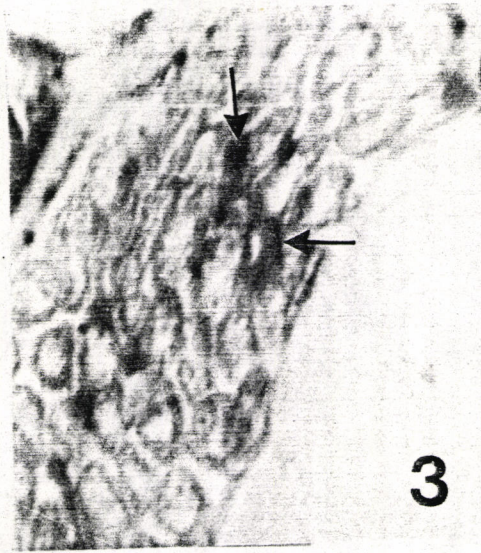


Fig. (3): Peripheral nerve, alpha naphthyl acetate esterase reaction in Schwann cells.

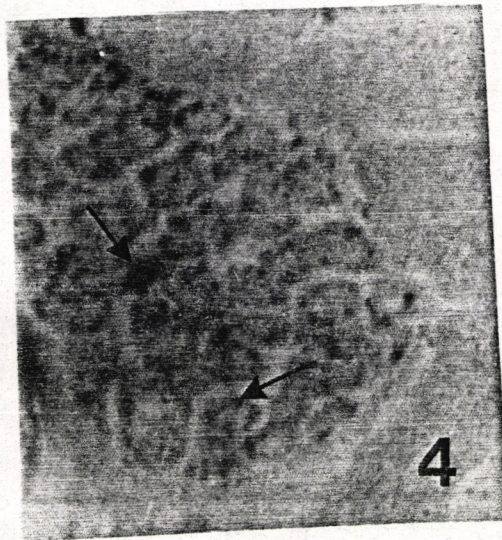


Fig. (4): Peripheral nerve, naphthol AS-D acetate esterase reaction.

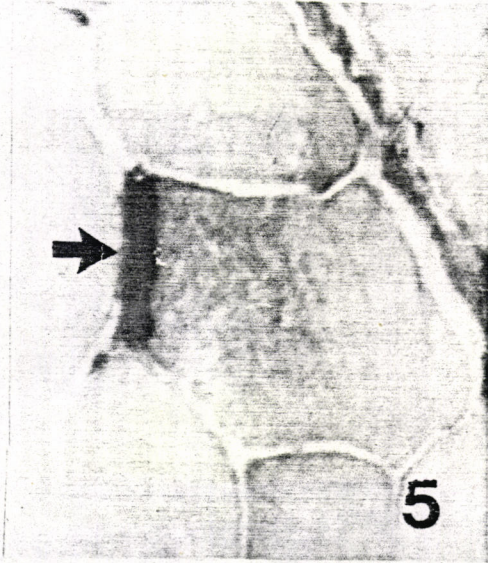


Fig. (5): alpha naphthyl acetate esterase reaction in the motor end-plate.

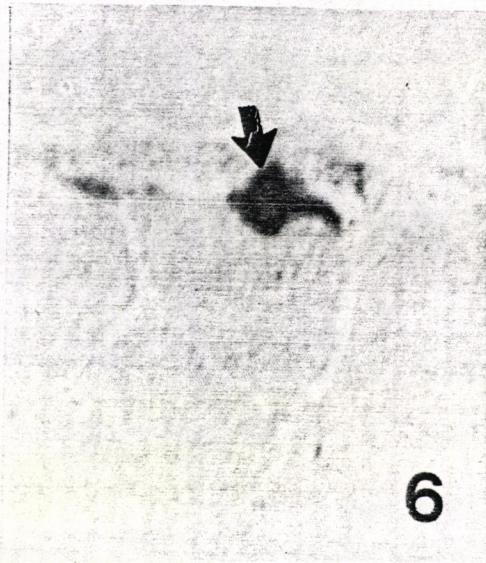


Fig. (6): Naphthol AS-D acetate esterase reaction in the motor and plate of EDL

This distribution is an indicator of a role played by these in the whole unit and not simply at the motor end-plate only.

Many papers have been published about the distribution of non-specific esterases and their variation in activity under endogenous and exogenous influences such as development, differentiation, nutrition, hormones, and diseases (7, 8 & 9).

At present time the metabolic function and the natural substrates of most non-specific esterases are still obscure. One can still only speculate about the functional importance. These speculations were made even more difficult by the fact that non-specific esterases are present as a complex tissue-specific mixture of various components, each of which presumably has a tissue-specific role. There is still a lack of biological interpretation of esterases in the nervous tissue despite the fact that it has been long since they were described in regions of the nervous system as the hypothalamus, neurohypophysis (10), and hippocampal region (1).

It seems that this particular distribution of esterases in the motor unit presents a tissue specific phenomenon, related to function, the intimate relations and influences of the motor neuron on the muscle fibres may be mediated through these non-specific esterases have a wide spectrum of catalytic activities, overlapping in their specificity for substrates particularly with some proteolytic enzymes especially endopeptidases.

Biological interpretation awaits a comprehension of the quantitative distribution of individual esterolytic enzymes. Therefore, the results of this study open the way for further evaluation of different nonspecific esterases (carboxylesterase, arylesterase, acetylerase... etc.) as markers for histochemical and physiological status of cells, tissues and organs, in particular the differentiation between various motor

neurons of the spinal cord and the influence of motor neurons on muscle fibres development and differentiation both in normal and diseased conditions.

REFERENCES

1. Iuppa, H. and Andra, J. (1983). The histochemistry of carboxyl ester hydrolases: Problems and possibilities. *Histochemical J.* 15:111-137.
2. Kasa, P. and Csillika, B. (1966). Electron microscopic localization of cholinesterase by a copper-Leas-thiicholine technique. *J. Neurochem.*, 13: 1345-1349.
3. Deimling O.V. and Bocling. A. (1976). Esterases in histochemistry and ultrahistochemistry. *Histochemical J.*, 8: 215-252.
4. Sharrad, W.J.W. (1955). The distribution of the permanent paralysis in the lower limb in poliomyelitis. *J. Bone and Joint Surgery*, 38:540-558.
5. Wary, S.H. (1966). Quoted by Gilliat, R.W. in: Axon branching in motor neurons, In: Andrew, B.L. (Editor) *Control and innervation of skeletal muscle*: pp. 53-63/ The University of St. Andrews.
6. Pearse, A.G.E. (1972). Carboxylic ester hydrolases. In: *Histochemistry, theoretical and applied* Vol. 2, 3rd. ed., pp. 761-807. Churchill Livingstone, Edinburgh, London.
7. Schiff, R. (1975). Esterase isoenzymes as markers in normal and disease processes, In: Markert, C. (Editor), *Isoenzymes*, pp. 775-797. Academic Press, New York.
8. Coates, P.M.; Mestriner, M.A. and Hopkinson, D.A. (1976). A preliminary genetic interpretation of esterase isoenzymes of human tissues, *Ann. Genet.*, 39: 1-20.

9. Kass, L. (1979). Cytochemistry of esterases. *SRO Crit. Rev. clin. lab.Sci.* 10: 295-223.
10. Pearse, A.G.E. (1958). Esterases of the hypothalamus and their functional significance. In: *pathiysiologica Diencephalica*, pp. 329-335. Springer, Vienna.

المجلة الطبية البيطرية العراقية المجلد الرابع عشر سنة (١٩٩٠)
دراسة كيميائية نسجية لانظيم الاستراز غير المحدد في الوصف
الحركة للارنب

انعم رشيد الصالحى، هاني طه العزاوي، هدى مهدي الخطيب
قسم التشريح - كلية الطب - جامعة بغداد

الخلاصة

تم دراسة انتشار فعالية انزيم استرازاسيتات ارليد حامض
النفثول الدالي في العصبونات المحركة في الشدق القطني
الاربع السفلية ثم الشدقة العجزية الاولى للنخاع الشوكي في
الارنب والتي يفترض انها قد تشترك في تزويد الباسطة الطويلة
لاباخص الطرف الخلفي في هذا الحيوان. كما تمت وفي عين الوقت
الدراسة نفسها على العصب المزود لهذه العضلة والمفأئح
الانتهائية المحركة في اليافها العظمية.
كانت الفعالية الاسترازية قوية في العصبونات المحركة في
النخاع الشوكي وفي خلايا شوان ومحاور الحزم العصبية
المحيطة والمفأئح الانتهائية المحركة في الالياف العظمية
لهذه العضلة. مما يسهل التعرف على هذه المفأئح.
تم مناقشة الاهمية البيولوجية لهذا الانزيم بصورة عامة
وفي المواقع المذكورة.