

Effect of selenium and melatonin on some parameters Related to metabolic syndrome induced by Acryl amide in male rats (Part I)

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Summary

This study was designed to investigate the ameliorative role of sodium selenite and melatonin on acryl amide induced metabolic syndrome in adult male rats. Twenty (20) adult male rats were randomly and equally divided into 4 groups (G1, G2, G3 and G4) and were treated orally for seven weeks. G1, (control group) was given distilled water, G2 were given orally 1mg/kg /B.W of Acryl amide, G3 group received 1mg/kg /B.W of Acryl amide followed by 0.5 mg/Kg/ B.W. of sodium selenite orally. G4 group received 1 mg/kg /B.W of acryl amide (orally) followed by 8 mg/Kg/ B.W. melatonin (IP/ injection). Fasting blood samples were collected by cardiac puncture at (0, 7 weeks) of the experiment for measuring serum total cholesterol, triacylglycerol, high density lipoprotein-cholesterol and serum uric acid, in addition to measuring waist circumference. The results showed an occurrence of central obesity, hyper uricemia, dyslipidemia (hyper cholesterolemia, triacyleglycerolemia and lowered high density lipoprotein-cholesterol concentration) in acryl amide treated animals. The results also indicated the alleviation of the changes in the above- mentioned parameters related to metabolic syndrome by sodium selenite and melatonin through lowering central obesity, total cholesterol, triacylglycerol, elevation of serum high density lipoprotein-cholesterol and lowering in serum uric acid.

Keywords: Acryl amide, Metabolic syndrome, Melatonin, Sodium selenite, Dyslipidemia.

Introduction

Metabolic syndrome (MetS) is a state of chronic low grade inflammation as a consequence of complex interplay between genetic and environmental factors, central obesity, atherogenic dyslipidemia, hyperuricemia, insulin resistance and hypertension are major criteria of the syndrome (1 and 2). Acryl amide (ACR) is produced in starchy foods those are baked, roasted or fried at high temperature (3). Bread, crisps, coffee and fried potato are the most contaminated food with ACR (4). The primary source of human exposure to acryl amide is occupational; other sources include food, drinking water, and smoking (5). Acryl amide is neurotoxic to experimental animal and human (6) and has mutagenic and carcinogen effect (7). Selenium, which is nutritionally essential for humans, is a constituent of more than two dozen selenoproteins that play critical roles in reproduction, DNA synthesis, and protection from oxidative damage and infection (8). Depending on the fact pointed to the role of reactive oxygen species and oxidative stress in the pathogenesis of may diseased condition correlated with metabolic

syndrome, this study aimed to evaluate the effect of sodium selenite and melatonin (the well-known antioxidants) on some parameters related to metabolic syndrome induced by Acryl amide in adult male rats.

Materials and Methods

Twenty adult male rats were randomly and equally divided into 4 groups (5/group) and treated orally (using gavage needle) for 7 weeks: G1: Rats of this group received distilled water and served as control, G2: Rats of this group received 1mg/kg B.W of Acryl amide, G3: Animals in the group received in addition to Acryl amide, 0.5 mg/Kg B.W. of sodium selenite, G4: Rats in this group received in addition to ACR, 8mg/Kg B.W. melatonin by interaperitoneal injection (IP). Fasting blood samples were collected at (0 and 7 weeks) of the experiment. Blood was drawn by cardiac puncture. Then serum samples were separated and frozen at -20°C until analysis of the following parameters: Metabolic syndrome parameters: A-Waist circumference using tape measure according to savory *et al.*, (9). B-Serum total cholesterol concentration using total cholesterol enzymatic kitlinear chemical,

(Barcelona, Spain) according to (10). D-Serum triacylglycerol concentration using TAG -C enzymatic kit (Merck company, U.K.) according to Fossati *et al* (11). E- Serum HDL-C concentration using HDL-C enzymatic kit (Bio system, Spain) according to Burstein *et al* (12). F- Serum uric acid concentration using uric enzymatic kit linear chemical, Barcelona (Spain) according to Fossati *et al* (13). Statistical analysis of data was performed on the basis of Two-Way Analysis of Variance (ANOVA) using a significant level of (P<0.05). Specific group differences were determined using least significant differences (LSD) as described by (14).

Results and Discussion

A Significant (P<0.05) reduction in waist circumference was observed in both groups which received sodium selenite (G3) or melatonin (G4) concurrently with ACR (G4) at the 3rd week of the experiment as compared with the values in ACR (G2) group (Table, 1). The results also showed that oral administration of sodium selenite or (I/P) melatonin concurrently with ACR caused

gradual significant reduction (P<0.05) in waist circumference in groups G3, G4 along the last four weeks (4th, 5th, 6th and 7th) of the experiment compared to the values in ACR and control groups.

Herein study showed significant (P<0.05) increase in serum total cholesterol (Table, 2) and triacylglycerol (TAG) concentration (Table, 3) in rats received 1mg/kg B.W ACR for 7 weeks (G2) groups compared to the values in other treated groups and control. Oral intubation of sodium selenite (group G3) or melatonin (group G4) concurrently with acryl amide caused significant elevation in serum HDL-C concentration compared to the values in ACR (G2) treated groups (Table, 4). At the end of the experiment, significant (P<0.05) increase in serum uric acid concentration was observed in Acryl amide (G2) treated group compared to the values in the control, G3 and G4 groups. On the other hand significant (P<0.05) decrease in serum uric acid concentration was observed after oral intubation of sodium selenite, or IP/injection of melatonin compared to the values in ACR treated group (Table, 5).

Table, 1: Effect of sodium selenite and melatonin on waist circumference (cm) in acryl amide treated rats.

Groups Periods (Weeks)	G1 Control	G2 Acrylamide (1mg/kgB.W)	G3 Acrylamide(1mg/kg B.W) and sodium selenite (0.5mg/kg B.W)	G4 Acrylamide (1mg/kgB.W) and melatonin (8mg/kg B.W)
1	14.71 ± 0.18 A C	14.78 ± 0.26 A c	14.71 ± 0.18 A a	14.71 ± 0.18 A a
2	14.71 ± 0.18 A C	15.04 ± 0.29 A bc	14.50 ± 0.17 A bc	14.64 ± 0.17 A a
3	15.08 ± 0.18 A B b	15.54 ± 0.24 A ab	14.28 ± 0.16 B abc	14.41 ± 0.17 B ab
4	15.27 ± 0.18 A b	15.75 ± 0.27 A a	14.04 ± 0.17 B bcd	14.34 ± 0.18 B ab
5	15.55 ± 0.19 A b	15.95 ± 0.28 A a	13.78 ± 0.18 B cd	14.20 ± 0.18 B cd
6	15.90 ± 0.20 A a	15.95 ± 0.28 A a	13.78 ± 0.18 B cd	14.20 ± 0.18 B ab
7	15.55 ± 0.20 A a	16.35 ± 0.27 B ab	13.45 ± 0.18 C d	13.91 ± 0.18 C b

Values are expressed as mean ±SE, n=5. Different small letters represent significant difference within group (P<0.05) vs. zero time. Different capital letters represent a significant difference between groups (P<0.05) vs. control.

Table, 2: Effect of sodium selenite and melatonin on serum total cholesterol (TC) concentration (mg/dl) in acrylamide treated rats.

Groups Periods (Weeks)	G1 Control	G2 Acryl amide (1mg/Kg B.W)	G3 Acrylamide(1mg/Kg B.W) and sodium selenite (0.5 mg/Kg B.W)	G4 Acrylamide (1mg/Kg B.W) and melatonin (8mg/Kg B.W)
Zero	77.39 ± 1.40 A a	79.77 ± 0.93 A b	75.92 ± 0.73 A b	76.35 ± 0.67 A b
7	78.73 ± 1.43 D a	128.49 ± 2.54 A a	119.85 ± 0.73 B a	99.32 ± 5.33 C a

Values are expressed as mean ±SE, n=5. Different small letters represent significant difference within group (P<0.05) vs. zero time. Different capital letters represent a significant difference between groups (P<0.05) vs. control.

Table, 3: Effect of sodium selenite and melatonin on serum triacylglycerol (TAG) concentration (mg/dl) in acrylamide treated rats.

Groups Periods (weeks)	G1 Control	G2 Acrylamide (1mg/Kg B.W)	G3 Acrylamide(1mg/KgB.W) and sodium selenite (0.5 mg/Kg B.W)	G4 Acrylamide (1mg/KgB.W) and melatonin (8mg/Kg B.W)
Zero	152.91 ±1.46 A a	155.16 ±1.61 A b	153.13 ±1.61 A b	156.96 ±2.02 A b
7	152.14 ± 1.40 D a	225.22 ±2.31 A a	199.08 ±2.01 B a	190.69 ± 3.02 C a

Values are expressed as mean ±SE, n=5. Different small letters represent significant difference within group (P<0.05) vs. zero time. Different capital letters represent a significant difference between groups (P<0.05) vs. control.

Table, 4: Effect of sodium selenite and melatonin on serum high density lipoprotein (HDL-C) concentration (mg/dl) in acrylamide treated rats.

Groups Periods (weeks)	G1 Control	G2 Acrylamide 1mg/Kg	G3 Acrylamide(1mg/Kg B.W) and sodium selenite(0.5mg/Kg B.W)	G4 Acrylamide (1mg/KgB.W) and melatonin (8mg/KgB.W)
Zero	47.39 ± 0.00 A a	44.80 ±1.55 B a	45.29 ± 0.74 AB a	44.68 ± 0.83 B a
7	45.72 ± 0.47 A a	23.46 ±1.48 C b	34.57 ± 1.10 B b	32.45 ± 0.71 B b

Values are expressed as mean ±SE, n=5. Different small letters represent significant difference within group (P<0.05) vs. zero time. Different capital letters represent a significant difference between groups (P<0.05) vs. control.

Table, 5: Effect of sodium selenite on serum uric acid concentration (mg/dl) in acrylamide treated rats.

Groups periods (weeks)	G1 Control	G2 Acrylamide (1mg/Kg B.W)	G3 Acrylamide(1mg/KgB.W) and sodium selenite (0.5mg/Kg B.W)	G4 Acrylamide (1mg/KgB.W) and melatonin (8mg/Kg B.W)
Zero	1.47 ± 0.15 A a	1.41 ± 0.04 A b	1.46 ± 0.10 A b	1.43 ± 0.11 A b
7	1.50 ± 0.10 C a	3.41 ± 0.10 A a	2.50 ± 0.04 B a	2.36 ± 0.25 B a

Values are expressed as mean ±SE, n=5. Different small letters represent significant difference within group (P<0.05) vs. zero time. Different capital letters represent a significant difference between groups (P<0.05) vs. control.

Similar results concerning dyslipidemia (elevation in TAG, TC and decrease in HDL-C) accompanied ACR treatment was observed by (15 and 16). Such results could be

attributed to the increment in the synthesis of plasma lipoprotein and high mobilization of lipid from liver (17). Insulin resistance induced after ACR exposure (18 and 19) may

lead to atherogenic dyslipidemia through suppression of lipolysis in adipocyte due to impairment in insulin signaling (20), elevated levels of free fatty acids (FFA) in the liver that serves as a substrate for the synthesis of TAG leading to triacylglycerolemia. Visceral obesity, recognized by increased waist circumference, is considered to be the first sign associated with metabolic syndrome (21). Visceral fat appears to be the most detrimental contributor to development of lipotoxicity in peripheral tissue by adipocytokines secretion (22). Adipocytokines secretion integrate autocrine and paracrine signal that mediate insulin sensitivity (23) and oxidative stress (24). The two criteria contribute to ACR toxicity and may be claimed to be responsible for visceral obesity accompanied ACR. On the other hand, positive correlation was found between adiponectin and HDL-C level (25), therefore, it can be concluded that IR accompanied ACR treatment may lead to decrease in adiponectin which coincides with the increase in visceral obesity as well as depression in HDL-C concentration.

It is speculated that in situations where free radical levels are high (e.g., obesity-related oxidant stress), Selenium supplementation will reduce free radical production and serum lipid composition (26), and thus visceral obesity. Selenium supplementation caused an increase in low-density lipoprotein (LDL) receptor activity and decrease the 3-OH-methyl-glutaryl CoA reductase expression (27) leading to decreased plasma LDL cholesterol and total cholesterol levels (28). This may be explaining the hypolipidemic effect of selenium. The result also showed significant increase in serum HDL-C concentration after sodium selenite supplementation indicating protective role against cardiovascular disease. Sodium selenite supplementation significantly increased the protein and mRNA expressions of apoA-I causing to elevation in HDL-C concentration (29).

The postulated anti-obesogenic effect of melatonin is, in part, a result of its regulatory role on the balance of energy, acting mainly on the regulation of the energy flux to and from the stores and in energy expenditure

(30). Additionally, it was demonstrated that even with an intact pineal production of melatonin, melatonin supplementation therapy in young animals reduces the size of the visceral fat deposits (by 50%) (31). Long-term administration of melatonin to humans and experimental animals reduces blood and liver cholesterol and LDL-cholesterol levels (32) and increased HDL-cholesterol in peri- and postmenopausal women (33). It has been proved that melatonin presence in the gall bladder enhanced conversion of cholesterol to bile, preventing oxidative stress causing hypocholesterolemia (34). Melatonin also decreases the amount of cholesterol produced in the gall bladder by regulating the cholesterol that passes through the intestinal wall.

Data regarding the effect of ACR on serum uric acid concentration (Hyperuricemia) in the current study is in accordance with other studies (19 and 35). A case of dyslipidemia referred to hypercholesterolemia, elevation in serum TAG and reduction in HDL-C concentration was found to be correlated with hyperuricemia and metabolic syndrome by some investigators (36). In the current study serum lipid profile was similarly affected by ACR which may explain its mechanism in hyperuricemia. Significant elevation in the reactive oxygen species and lipid peroxidation inducing oxidative stress by ACR accompanied by depletion in the antioxidant level of kidney (37) could impair renal function leading to hyperuricemia.

Sodium selenite supplementation caused reduction in serum creatinine, urea and uric acid in rats exposed to acryl amide suggesting the protective effect of selenium on renal function (35 and 38). The antioxidant properties of selenium could be attributed to its renoprotective effect. Oxidative stress induced complications of diabetes may include stroke, neuropathy, retinopathy and nephropathy (39) and thus improving oxidative stress by selenium will lead to renoprotection and decrease in uric acid. Hyperuricemia induced after melatonin treatment could be attributed to its antioxidant effect that restored renal function. Melatonin stimulates synthesis of antioxidant enzymes, increased activity of others and protect

antioxidant enzyme from damage (40 and 41). Serum creatinine and urea nitrogen concentration was significantly restored to their normal values after co-treatment with melatonin in different types of nephrotoxicity (42) indicating its renoprotective effect.

References

1. Spolidoro, J. V. (2013). Waist circumference in children and adolescents correlate with metabolic syndrome and fat deposits in young adults. *Clin. Nutr.*, 32(1):93-97.
2. Kaur, J. (2014). A comprehensive review on metabolic syndrome. *Cardiol. Res. Pract.*, Volume 2014. ID943162. 21 Pages.
3. Rommens, C. M.; Yan, H.; Swords, K.; Richael, C. and Ye, J. (2008). Low acrylamide French fries and potatochips. *Plant Biotechnol. J.*, 6(8):843–853.
4. Freisling, H.; Moskal, A.; Ferrari, P.; Nicolas, G.; Knaze, V. and Clavel-Chapelon, F. (2013). Dietary acrylamide intake of adults in the European Prospective Investigation into Cancer and Nutrition differs greatly according to geographical region. *Europ. J. Nutr.*, 52(4):1369-1380.
5. Allam, A.; Abdul-Hamid, M.; Bakery, A. E.; El-Ghareeb, A.W.; Ajarem, J. S. and Sabri, M. (2013). Effect of acrylamide on cerebral neuron developments in albino rats. *Life Sci J.*, 10(3):1760-1771.
6. Zhang, L.; Gavin, T.; Barber, D. S. and LoPachin, R. M. (2011). Role of the Nrf2-ARE pathway in acrylamide neurotoxicity. *Toxicol. Lett.*, 205(1):1–7.
7. DeWoskin, R. D.; Sweeney, L. M.; Teegarden, J. G.; Sams, R. and Vandenberg, J. (2013). Comparison of PBTK model and biomarker based estimates of the internal dosimetry of acrylamide. *Food Chem. Toxicol.*, 58:506-521.
8. Sadeghian, S.; Kojouri, G. A. and Mohebbi, M. (2012). Nanoparticles of selenium as species with stronger physiological effects in sheep in comparison with sodium selenite. *Biol. Trace Elem. Res.*, 146(3):302-308.
9. Savory, L. A.; Ker, C. J.; Whitin, P.; Fine, N.; McEnen, J. and Ashton, T. (2012). Selenium supplementation and exercise: Effect on oxidant stress in overweight adults. *Obesity Res.*, J., 20(4):794–801.
10. Allain, C. C.; Poon, L. S.; Chon, C. S.; Richmond, W. and Fu, P. C. (1974). Enzymatic determination of total serum cholesterol, *Clin. Chem.*, 20:470-475.
11. Fossati, P. and Prencipe, L. (1982). Serum triglycerides determined colorimetrically with an enzyme that produces hydrogen peroxide. *Clin. Chem.*, 28:2077-2080.
12. Burstein, M.; Scholnick, H. R. and Morfin, R. (1980). Rapid method for the isolation of lipoproteins from human serum by precipitation with polyanions. *Scand. J. Clin. Lab. Invest.*, 40:583-595.
13. Fossati, P.; Prencipe, L. and Berti, G. (1980). Use of 3,5-dichloro -2- hydroxybenzene sulfonic acid/ 4-aminophenazone chromogenic system in direct enzymic assay of uric acid in serum and urine. *Clin. Chem.*, 26:227-231.
14. Snedecor, G. W. and Cochran, W. G. (1973). *Statistical Methods*. 6th the Iowa state University Press., Pp: 238-248.
15. El -Shafey, A. A. M. S. M.; Nour El -Din, E. M.; Mohamed. S. M. M. and Ezaat, A. M. A. E. (2013). Protective effect of garlic "Allium sativum" and karkada "Hibiscus sabdarrifa" on acrylamide treated male albino rats. *Egypt. J. Exp. Biol. (Zool.)*, 9(1): 101-107.
16. Eid, M. M.; Ahmed, A. A.; Saleh, N. T. and Sheir, M. A. (2012). Effect of some phytochemicals on reduction of acrylamide in fried potato chips and their biological effect on blood lipid profile. *Minia J. Agric. Res. and Develop.*, 32(6):939-963.
17. Rawi, S. R.; Marie, M.A.S.; Fahmy, S. R. and El-Abied, S. A. (2012). Hazardous effects of acrylamide on immature male and female rats. *Afr. J. Pharm. Pharmacol.*, 6(18):1367-1386.
18. Lin, C.Y.; Lin, Y.C.; Kuo, H. K.; Hwang, J. J.; Lin, J. L.; Chen, P. C. and Lin, L.Y. (2009). Association among acrylamide, blood insulin, and insulin resistance in adults. *Diabetes Care*, 32(12):2206-2211.
19. AL-Agele, F.A.L. (2014). Effect of acrylamide on some parameter related to metabolic syndrome relative to fructose in adult male rats. MSc. Thesis, University of Baghdad /College of Veterinary Medicine.
20. Totani, N.; Yawata, M.; Ojiri, Y. and Fujioka, Y. (2007). Effects of trace

- acrylamide intake in Wister rats. *J. Oleo. Sci.*, 56:501-506.
21. Grundy, S. M.; Hansen, B.; Smith, S. C.; Cleeman, J. I. and Kahn, R. A. (2004). Clinical management of metabolic syndrome: Report of the american heart association/national heart, lung, and blood Institute/American Diabetes Association Conference on scientific issues related to management. *Circulation*. 109(4):551–556
 22. Gill, H.; Mugo, M.; Whaley-Connell, A.; Stump, C. and Sowers, J. (2005). The key role of insulin resistance in the cardiometabolic syndrome. *Am. J. Med. Sci.*, 330(6):290-294.
 23. Saleem, U.; Khaleghi, M.; Morgenthaler, N. G.; Bergmann, A.; Struck, J.; Mosley, T. H. and Kullo, I. J. (2009). Plasma carboxy-terminal provasopressin (copeptin): a novel marker of insulin resistance and metabolic syndrome. *J. Clin. Endocrinol. Metabol.*, 94(7):2558–2564.
 24. Tsimikas, S.; Willeit, J.; Knoflach, M.; Mayr, M.; Egger, G.; Notdurfter, M.; Witztum, J. L.; Wiedemann, C. J.; Xu, Q. and Kiechl, S. (2009). Lipoprotein-associated phospholipase A2 activity, ferritin levels, metabolic syndrome, and 10-year cardiovascular and non-cardiovascular mortality: results from the Bruneck study. *European Heart J.*, 30(1):107-115.
 25. Fain, J. N.; Cheema, P. S.; Bahouth, S. W. and Hiler, M. L. (2003). Resistin release by human adipose tissue explants in primary culture. *Biochem. Biophys. Res. Comm.*, 300(3):674–678.
 26. Alissa, E. M.; Bahijri, S. M. and Ferns, G. A. (2003). The controversy surrounding selenium and cardiovascular disease: a review of the evidence. *Med. Sci. Monit.*, 9:RA9–RA18.
 27. Dhingra, S. and Bansal, M. P. (2006). Attenuation of LDL receptor gene expression by selenium deficiency during hypercholesterolemia. *Mol. Cell Biochem.*, 282(1-2):75-82.
 28. Yang, Y. R.; Meng, F. C.; Wang, P.; Jiang, Y. B.; Yin, Q.; Chang, J.; Zuo, R.Y.; Zeng, Q.H. and Liu, J.X. (2012). Effect of organic and inorganic selenium supplementation on growth performance, meat quality and antioxidant property of broilers. *Afr. J. Biotechnol.*, 11:3031-3036.
 29. Putri, M.; Yamazaki, C.; Syamsunarno, Mas Rizky, A. A.; Puspitasari, I. M.; Abdulah, R.; Kameo, S.; Iso, T.; Kurabayashi, M. and Koyama, H. (2014). Effect of sodium selenite supplementation on pre β -high-density lipoprotein formation-related proteins in human primary hepatocytes. *IJFANS*. 3(6):16-22.
 30. Bartness, T. J.; Demas, G. E. and Song, C. K. (2002). Seasonal changes in adiposity: the roles of the photoperiod, melatonin and other hormones, and sympathetic nervous syst. *Exp. Biol. Med.*, 227:363–376.
 31. Tan, D. X.; Manchester, L. C.; Fuentes-Broto, L.; Paredes, S. D. and Reiter, R. J. (2011). Significance and application of melatonin in the regulation of brown adipose tissue metabolism: relation to human obesity. *Obes. Rev.*, 12(3):167–188.
 32. Hussain, S. A. (2007). Effect of melatonin on cholesterol absorption in rats. *J. Pineal Res.*, 42:267- 271.
 33. Tamura, H.; Nakamura, Y.; Narimatsu, A.; Yamagata, Y.; Takasaki, A.; Reiter, R. J. and Sugino, N. (2008). Melatonin treatment in peri- and postmenopausal women elevates serum high-density lipoprotein cholesterol levels without influencing total cholesterol levels. *J. Pineal Res.*, 45:101-105.
 34. Koppiseti, S.; Jenigiri, B.; Terron, M. P.; Tengattini, S.; Tamura, H.; Flores, L. J.; Tan, D. X. and Reiter, R. J. (2008). Reactive oxygen species and the hypomotility of the gall bladder as targets for the treatment of gallstones with melatonin: a review. *Dig. Dis. Sci.*, 53(10):2592–2603.
 35. Teodor, V.; Cuciureanu, M.; Slencu, B. J.; Zamosteanu, N. and Cuciureanu, R. (2011). Potential protective role of selenium in acrylamide intoxication. A biochemical study. *Studia Universitatis “Vasile Goldiş”, Seria Stiințele Vietiivol.* 21(2):263-268.
 36. Khashab, M. A.; Liangpunsakul, S. and Chalasani, N. (2008). Nonalcoholic fatty liver disease as a component of the metabolic syndrome. *Curr. Gastroenterol. Rep.*, 10:73–80.
 37. ALTurfan, I. E.; Beceren, A.; Şehirli, O. A.; Demiralp, E. Z.; Şener, G. and Omurtag, Z. G. (2011). Protective effect of N-acetyl-L-

- cysteine against acrylamide-induced oxidative stress in rats. Turk. J. Vet. Anim. Sci., 36(4):438-445.
38. Ismail, S. N. (2014). Red ginseng extract and selenium reduce oxidative stress induced by potassium bromate in male rats. Alex. J. Agric. Res., 59(1):1-8.
39. Asmat, U.; Abad, K. and Ismail, K. (2015). Diabetes mellitus and oxidative stress-A concise (review). Saudi Pharmaceut J., Available Online. In Press.
40. Mayo, J. C.; Tan, D. X.; Sainz, R. M.; Lopez-Burillo, S. and Reiter, R. J. (2009). Oxidative damage to catalase induced by peroxy radical: Functional protection by melatonin and other antioxidants. Free Radic Res., 37:543-553.
41. Sener, G.; Paskaloglu, K.; Toklu, H.; Kapucu, C.; Ayanoglu-Dulger, G. and Kacmaz, A. (2004). Melatonin ameliorates chronic renal failure-induced oxidative organ damage in rats. J. Pineal Res., 36(4):232-241.
42. Kilic, U.; Kilic, E.; Tuzcu, Z.; Tuzcu, M.; Ozercan, I. H.; Yilmaz, O.; Sahin, F. and Sahin, K. (2013). Melatonin suppresses cisplatin-induced nephrotoxicity via activation of Nrf-2/HO-1 pathway. Nutr. Metab., 10(1):7.

تأثير السيلينيوم والميلاتونين في بعض المعايير المتعلقة بمتلازمة الأيض المستحدث بالاكريلاميد في ذكور الجرذان (المبحث الأول)

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الخلاصة

صُممت هذه الدراسة لمعرفة الدور التحسيني لسيلينيوم والصوديوم والميلاتونين في بعض المعايير المتعلقة بمتلازمة الأيض في ذكور الجرذان المستحدثة باستعمال الاكريلاميد. استعملت (20) من ذكور الجرذان البالغة قسمت عشوائياً على اربع مجاميع (4) متساوية وعملت لمدة (7) اسابيع: المجموعة الاولى (G1) اعطيت الماء مقطر وعدت مجموعة سيطرة، في حين جرعت جرذان المجموعة الثانية (G2) فموياً "1ملغم/كغم من وزن الجسم"، المجموعة الثالثة (G3) فضلاً عن الاكريلاميد (1ملغم/كغم من وزن الجسم)، 0.5 ملغم/كغم من وزن الجسم من سيلينيوم والصوديوم وأما حيوانات المجموعة الرابعة (G4) فقد حقنت عبر الغشاء البريتوني 8 ملغم/كغم من وزن الجسم من الميلاتونين بعد تجريعها فموياً 1ملغم/كغم من وزن الجسم. جمعت عينات الدم في بداية التجربة وبعد 7 اسابيع من التجربة بطريقة الوحز القلبي لغرض قياس تركيز الكولسترول الكلي والدهون الثلاثية، الكولسترول في الشحوم البروتينية عالية الكثافة وحامض اليوريك فضلاً عن قياس الخصر للدلالة على السمنة المركزية. اظهرت النتائج حدوث متلازمة الأيض بالاكريلاميد تمثلت بالسمنة المركزية واضطراب في ابيض الدهون من خلال ارتفاع في تركيز الكولسترول والدهون الثلاثية مع انخفاض تركيز الكولسترول في الشحوم البروتينية عالية الكثافة وارتفاع في تركيز حامض اليوريك. واطهرت النتائج أيضاً الدور الايجابي للسيلينيوم والصوديوم والميلاتونين عن تقليل حدة المعايير السابقة متمثلة بحدوث انخفاض في السمنة المركزية وتركيز الكولسترول والدهون الثلاثية وارتفاع في تركيز الكولسترول في الشحوم البروتينية عالية الكثافة مع انخفاض في تركيز حامض اليوريك.

الكلمات المفتاحية: الأكريلاميد، متلازمة التمثيل الغذائي، الميلاتونين، الصوديوم سيلينيوم، دسليبيديميا.